

WATER-SOLUBLE FULLERENE C₆₀(OH)₂₄ MODULATES GROWTH AND PROLIFERATION OF K562 HUMAN ERYTHROLEUKEMIA CELL LINE

DIMITAR JAKIMOV¹, GORDANA BOGDANOVIĆ¹, MIRA BALTIĆ¹, SLAVICA TURŠIĆ¹,
LIDIJA ALEKSIĆ¹, ALEKSANDAR ĐORĐEVIĆ², JASMINKA MRĐANOVIĆ¹, MIRJANA
VOJINOVIĆ-MILORADOV²

¹ Experimental Oncology Department and Laboratory for Immunobiology,
Institute of Oncology Sremska Kamenica,
Institutski put 4, 21204 Sremska Kamenica
YUGOSLAVIA

² Department of Chemistry,
Faculty of Science, University of Novi Sad,
Trg Dositeja Obradovića 3, 21000 Novi Sad
YUGOSLAVIA

Abstract: - Fullerenes and their water-soluble derivatives have recently become a topic of interest in biomedicine. Here we report on growth and proliferation activity of fullereneol C₆₀(OH)₂₄ on K562 human erythroleukemia cells. Capability of K562 cells to synthesize a DNA by incorporation of radiolabeled thymidine, their mitotic activity and cell cycle distribution has been evaluated 3, 24 and 48 hours after fullereneol treatment. Viability of fullereneol treated K562 cells was up to 5% lower in all samples in comparison to control, regardless C₆₀(OH)₂₄ concentration or incubation period of cells with fullereneol. No significant differences were found in morphological appearance between control and fullereneol treated cells analyzed by light microscopy. A mitotic index of fullereneol treated cells was decreased in comparison to control regardless the period of cell incubation with fullereneol. Synthesis of DNA of fullerene treated K562 cells was inhibited, but only at the highest fullereneol concentration. The cell cycle distribution pattern of K562 cells treated with fullereneol at concentration of 0.041 μmol/l was comparable to that of the control but increasing number of cells in G₂M phase was observed during the treatment period.

Water-soluble [C₆₀] fullerene derivative, C₆₀(OH)₂₄, modulates cell cycle of the K562 cell influencing both synthetic and mitotic cell cycle phase

Key-Words: - Fullerene; Cell line; Cell cycle; DNA synthesis; Mitosis.

1 Introduction

Fullerenes are relatively new group of compounds and represent a class of sphere-shaped molecules made exclusively of carbon atoms. Since they were discovered in 1985 [1], many aspects of both fullerene [C₆₀] and its analogous have been intensively studied to reveal their physical and chemical reactivity [2-7]. However, mainly due to high hydrophobicity and poor solubility of the compounds in aqueous media, their biological activities have not been fully investigated.

The chemical modification of fullerene [C₆₀] molecule toward water-soluble adducts resulted in compounds exhibiting a variety of biological activities [2,3,6,8-10]. [C₆₀] fullerene derivatives cleave double-strand DNA upon visible light irradiation [7,9,11]. Cytotoxicity to various tumor

cell lines [9-10] and distinct inhibiting activities against various enzymes (some proteases and reverse transcriptases) were also reported [12-14]. Anti-apoptotic, anti-bacterial and virocidal activity of [C₆₀] derivative, as well as neuroprotective effects of polyhydroxylated [C₆₀] fullerene derivatives and polar carboxy acid [C₆₀] derivatives were reported [15-20]. Several *in vitro* and *in vivo* studies have shown that fullereneols, polyhydroxylated [C₆₀] fullerene derivatives, are free radical scavengers and potent antioxidative agents in biological systems [15,17,21-24]. Antiproliferative effect of fullereneols, as well as their enzyme inhibition activity, can be attributed to their antioxidative property [3,15,25-28].

Our preliminary results showed that fullereneol C₆₀(OH)₂₄, continuously present in culture up to 48

hours, at nanomole concentrations, induced low but transient growth inhibition of several human tumor cell lines [10]. A mechanism of growth-inhibitory activity of the fullereneol might be explained by inhibition of mitotic spindle formation [34].

The aim of this study was to investigate the influence of nanomolar concentrations of fullereneol $C_{60}(OH)_{24}$ on growth and proliferation activity of K562 cells estimating their synthetic and mitotic cell cycle phase.

2 Materials and Methods

2.1 Cell line

The human erythroleukemia cell line K562 grows in RPMI 1640 medium (Sigma) supplemented with 2 mM glutamine, 10% FCS (fetal calf serum, NIVNS) and antibiotics: 100 IU/ml of penicillin and 100 μ g/ml of streptomycin (ICN). The cells were sub-cultured twice a week at the concentration of 50,000-100,000 cell/ml. Cells were cultured in flasks (Costar, 25 cm²) and incubated at 37 °C in the 100% humidity atmosphere and 5% of CO₂.

2.2 Fullerene derivative

Polyhydroxylated fullerene derivative, fullereneol $C_{60}(OH)_{24}$, was synthesized and characterized in the laboratories of the Chemistry Department, Faculty of Sciences, University of Novi Sad. The fullereneol $C_{60}(OH)_{24}$ was yielded by complete substitution of bromine atoms from $C_{60}Br_{24}$ with hydroxyl groups [4-5].

The fullereneol was dissolved in 1 ml of distilled water ("stock" solution). Dissolved substance was further diluted with RPMI 1640 medium to set an array of "working" solutions of proper concentration, which were added in volume of 10 μ l/culture to achieve final concentrations of 0.04, 0.18 and 0.71 μ mol/l.

2.3 Cell treatment

Samples of control and treated K562 cells were seeded in 10ml tubes (10⁵ cells/ 4 ml of RPMI 1640 medium with 10% FCS) for mitotic index determination or in 96-well microtiter plates (1x10⁵ cells /180 μ l RPMI 1640 medium with 10 % of FCS) for DNA synthesis evaluation. Cells were incubated at 37 °C in the 100% humidity atmosphere and 5% of CO₂, for 3, 24 and 48h. Three different concentrations of the fullereneol were added into culture medium: 0.04, 0.18 and 0.71 μ mol/l.

2.4 Viability and cell density

Population cell density (a number of cells per volume unit) and percentage of viable cells were performed by DET (dye exclusion test) as previously described [35].

2.5 Determination of mitotic index

Slide smears (cytocentrifuge preparations) were prepared from both control and fullereneol treated cells (3x10⁴ cells / 100 μ l from each sample in duplicate) on the cytopspin centrifuge (Shandon). Staining of dried specimens were performed by May Grunwald - Giemsa method (MGG). The slides were observed under the light microscope with 400x magnification. At least 500 cells were counted on each specimen, within five different view fields per specimen, and at least two specimens were observed for each sample.

Cells were counted in the mitosis (all phases) and mitotic index (MI) was estimated as follows:

$$MI = [\text{No. of cells in mitosis}] / [\text{total No. of cells}] \times 100$$

2.6 ³H-Thymidine incorporation assay

Aqueous solution of ³H-Thymidine (³H-Thy) (specific activity of 185 GBq/mmol, Amersham) was diluted with RPMI 1640 culture medium in ratio 1:20. Two hours before the cells harvesting, 20 μ l of diluted ³H-Thy was added in every well of the plates (1 μ Ci / well). After the incubation, the K562 cells were harvested on filter-paper disks (Scatron Titertek), dried overnight at 40°C, transferred into vials with 5 ml of scintillation liquid and counted in the liquid scintillation beta-counter (TM Analytic BetaTrac 6895). Radioactivity of the samples was expressed as DPM (Disintegrations Per Minute) and is proportional to the amount of ³H-Thy incorporated into DNA of cultured cells. Results were presented as a proliferation index i.e. DPM ratio of treated and control samples.

2.7 Cell cycle analysis by flow cytometry

Cell suspension (1x10⁶) was treated with 1ml of 0.1% Triton-x-100 for five minutes at 4°C, followed by 20 μ l RNA-ase (1mg/ml) in PBS and stained with propidium iodide (PI). The cellular DNA content was measured using FACScan flow cytometer equipped with CellQuest software (Becton Dickinson).

3 Results

A viability of fullerene treated K562 cells was comparable to control (96.3-98.1) and remained high regardless of fullerene concentration or incubation period of cells with fullerene (92.6 - 99.1). Small (by 10 to 14 %) but persistent growth inhibition of fulleranol, at concentration of 0.04 $\mu\text{mol/l}$, on K562 cells was found. The same growth pattern was observed in 72-hour culture previously treated with fulleranol for 48 hours (Fig 1).

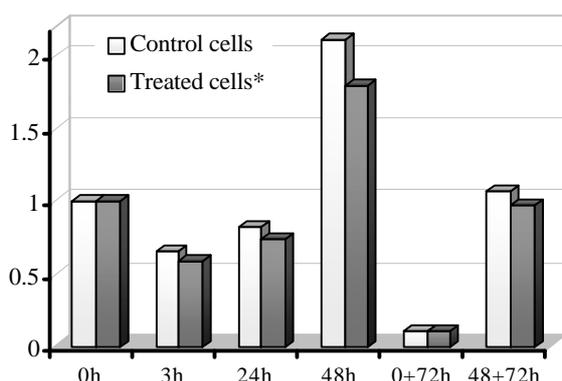


Fig. 1: Total number ($\times 10^6$) of K562 cells: untreated, fulleranol treated (0.04 $\mu\text{mol/l}$) during 3, 24 and 48 hours and 72 hours after 48-hour fulleranol treatment.

No significant differences were found in morphological appearance between control and fulleranol treated cells analyzed by light microscopy, concerning cell size, shape and cytoplasm/nucleus ratio.

A mitotic index of fulleranol treated (0.04 $\mu\text{mol/l}$) cells was decreased during the incubation period up to 30 percent in comparison to control (Table 1).

Table 1: Mitotic index of K562 cells.

Fulleranol conc. ($\mu\text{mol/l}$)	Mitotic index (%)		
	3h	24h	48h
0	3.85%	3.77%	3.71%
0.4	2.70%	2.76%	2.36%

K₅₆₂ cells were treated for 3, 24 or 48 hours with 0.04 $\mu\text{mol/l}$ of fulleranol C₆₀(OH)₂₄, Each value (MI) was obtained by counting at least 1000 cells per sample

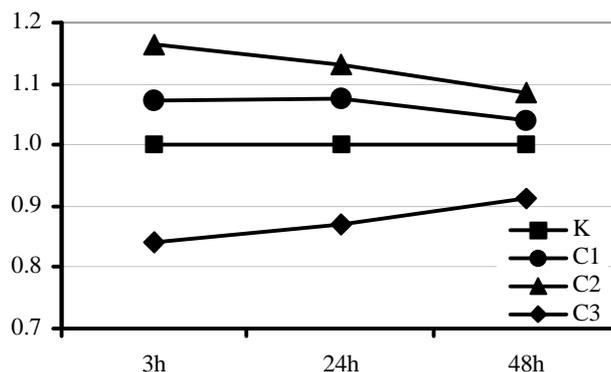


Fig. 2: DNA synthesis of K562 cells treated for 3, 24 or 48 hours with 0.04 (C1), 0.18 (C2) and 0.71(C3) $\mu\text{mol/l}$ of fulleranol C₆₀(OH)₂₄. Each value (proliferation index) is the mean of six replicates.

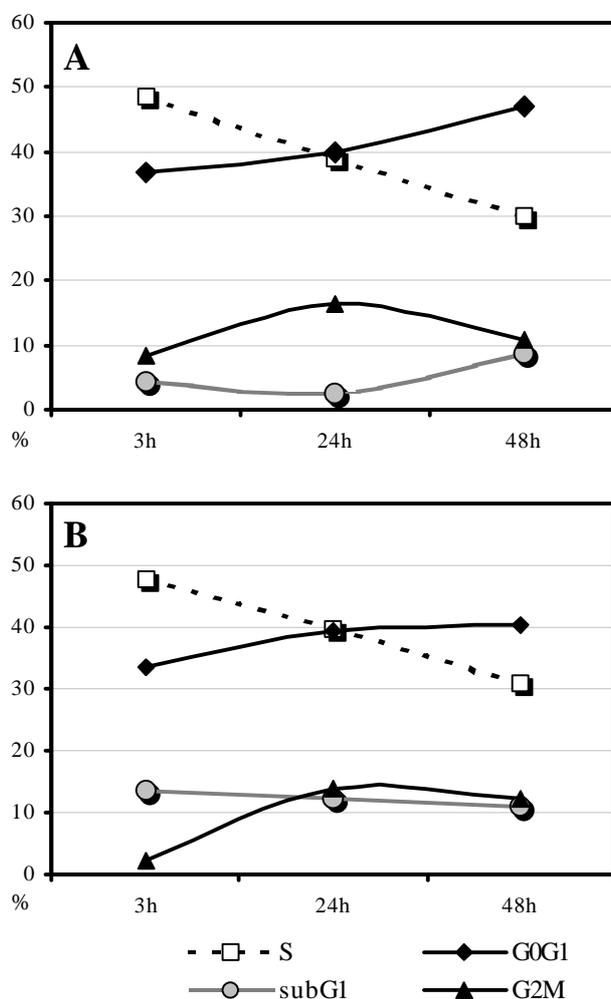


Fig. 3: Flow cytometry analysis of the cell cycle distribution of K562 cells after 3, 24 or 48 hours of culture. A = control K562 cells; B = fulleranol (0.04 $\mu\text{mol/l}$) treated K562 cells.

DNA synthesis of fullereneol treated K562 cells was also inhibited but only at the highest fullereneol concentration (Fig. 2).

The cell cycle distribution pattern of K562 cells treated with fullereneol at concentration of 0.041 μmol/l was comparable to the control but increasing number of cells in G2M phase was observed during the treatment period. The proportion of sub-G1 cells was higher in comparison to control (11-13 vs. 3-8%) (Fig. 3).

4 Discussion

The results from previous studies on polyhydroxylated fullerene [C₆₀] derivative activity in human tumor cell lines showed that fullereneol C₆₀(OH)₂₄ induced low but transient cell growth inhibition during 48-hour cell treatment [10].

This study confirmed low growth-inhibitory activity of fullereneol C₆₀(OH)₂₄ on K562 cell. High cell viability (above 92%) of all fullereneol treated samples additionally confirms low cytotoxicity of water-soluble [C₆₀] derivative. Synthetic and mitotic cell cycle phases of K562 cells were evaluated by determination of the mitotic index and the index of proliferation. Decreased mitotic index of treated cells in relation to control ones (i.e. smaller number of cells in various mitotic phases during 48 hours), as well as changes in DNA synthesis indicates that fullereneol modulates cell cycle of K562 cells.

The finding of decreased MI supports previously mentioned theory that C₆₀(OH)₂₄ inhibits the formation of mitotic spindle and microtubule assembly during cell division [34].

DNA synthesis of K562 cells was inhibited - proliferation index is lower for each time point but only for the highest fullereneol concentration.

The cell cycle analysis using flow cytometry with PI showed that cell cycle distribution pattern of K562 cells treated with nanomolar fullereneol concentration, was similar to that of control. However, increasing number of cells in G2M and G1 phases were observed during the treatment period being almost unchanged during last 24 hours of culture. In fullereneol treated cells a proportion of sub-G1 cells were remained during treatment period indicating that fullereneol induced apoptosis in K562 cells. To verify this last observation it would be necessary to analyze both the cell cycle and apoptosis induction using a range of fullereneol concentration and some other methods for apoptosis measurement.

Recently, some studies reported on antiproliferative effects of fullereneols containing 18-20 OH-groups

per C₆₀ molecule on normal vascular smooth muscle cells and tumor cell lines as well.^{4,20} The results related to the study of Nakamura et al. [9] can be explained by interaction of ¹O₂, (generated upon light excitation of fullerene), with cells or by direct interaction between the fullerene excited state and cells. In the second study inhibitory effect on cell growth was explained by the fullereneol antioxidative properties and partly by inhibition of signal transduction through tyrosine kinase pathway [10].

The persistent decrease of cell number might be a result of the fullereneol that entered the cells and induced growth inhibition by mechanism unexplained so far. It is already confirmed, by *in vitro* and *in vivo* studies, that fullerenes can be taken up by cells and be accumulated in some organs, such as liver, bone tissue and hair.^{11,21} There are scarce data on their metabolism and their chronic effects in biological systems as well.¹¹ Small but sustained growth inhibition of K562 cells even 72 hours after 48 hour fullereneol treatment is in accordance with the assumption that it is withheld by the cells.

The results obtained - small but persistent decrease in cell number with high cell viability, small changes in cell cycle distribution, DNA synthesis and mitotic activity indicate that fullereneol C₆₀(OH)₂₄ exerts a cytostatic rather than a cytotoxic activity against K562 cells.

The examinations of the biological properties of water-soluble fullerenes are still insufficient. Further investigations of their effects on other different cell line types and explanation of the mechanism of their action on cell level are needed.

5 Conclusion

Water-soluble [C₆₀] fullerene derivative, C₆₀(OH)₂₄, modulates cell cycle of the K562 cell influencing both synthetic and mitotic cell cycle phase. The results obtained indicate that fullereneol C₆₀(OH)₂₄ exerts a cytostatic rather than a cytotoxic activity against K562 cells.

Acknowledgment: - This work was supported by the grant No.1893 from the Ministry for Science, Technologies and Development of the Republic of Serbia.

References:

- [1] Kroto H. W., Heath J. R., O'Brien R. F., Curl R. F. and Smalley R. E., Buckminsterfullerene, *Nature*, Vol.318, 1988, pp. 162-163.

- [2] Vojinoviæ-Miloradov M. and Koruga Dj., Fullerene C₆₀ and its Applications in Biomedicine, *Archive of Oncology*, Vol.5, No.3, 1997, pp. 109-117.
- [3] Okuda K., Hirota T., Hirobe M., Nagano T., Mochizuki M. and Mashino T., Synthesis of Various Water-Soluble C₆₀ Derivatives and Their Superoxide-Quenching Activity, *Fullerene Science and Technology*, Vol.8, No.1&2, 2000, pp. 89-104.
- [4] Ðorðeviæ A., Vojinoviæ-Miloradov M., Adamov J.M., Lazar D., Èanadi J., Ribar B., Hlpka J., Ðorðeviæ-Miliæ V., Deveèerski A. and Popoviæ N., Synthesis of Fullerene Bromine Derivatives, *Fullerenes and Nanotubes Review*, Vol.1, No.2-3, 1997, pp. 86-87.
- [5] Ðorðeviæ A., *Synthesis and Characterization of Water-Soluble Fullerene Derivative - molecule C₆₀*, Thesis, University of Novi Sad, 2000.
- [6] Chi Yu, Jayandra B. Bhonsle, Lee Y. Wang, Jao G. Lin, Bao-Ji Chen and Long Y. Chiang, Synthetic aspects and Free-Radical Scavenging efficiency of Polyhydroxylated C₆₀, *Fullerene Science and Technology*, Vol.5, No.7, 1997, pp. 1407-1421.
- [7] Yamakoshi Y., Sueyoshi S., Fukuhara K., Miyata N. Masumizu T. and Kohno M., •OH and O₂^{•-} Generation in Aqueous C₆₀ and C₇₀ Solutions by Photoirradiation: An EPR Study, *Journal of the American Chemical Society*, Vol.120, No.47, 1996, pp. 12363-12364.
- [8] Jensen A. W., Wilson S. R. and Schuster D. I., Biological Applications of Fullerenes, *Bioorganic and Medicinal Chemistry*, Vol.4, No.6, 1996, pp. 767-779.
- [9] Nakamura E., Tokuyama H., Yamago S., Shiraki T. and Sugiura Y., Biological Activity of Water-Soluble Fullerenes. Structural Dependence of DNA Cleavage, Cytotoxicity, and Enzyme Inhibitory Activities Including HIV-Protease inhibition, *Bulletin of the Chemical Society of Japan*, Vol.69, No.8, 1996, pp. 2143-2151.
- [10] Bogdanoviæ G., Vojinoviæ-Miloradov M., Kojiæ V., Ðorðeviæ A., Èanadi J., Koruga Ð., Baltiæ V.V. and Tabš D., Biological Activity of Water-Soluble Fullerene: C₆₀(OH)₂₄, *Archive of Oncology*, Vol.5, No.3, 1997, pp. 147-149.
- [11] Yamakoshi Y., Yagami T., Sueyoshi S. and Miyata N., Acridine Adduct of [60] Fullerene with Enhanced DNA-Cleaving Activity, *The Journal of Organic Chemistry*, Vol.61, No.21, 1996, pp. 7236-7237
- [12] Satoh M., Matsuo K., Kiriya H., Mashino T., Hirobe M. and Takayanagi I., Inhibitory Effect of a Fullerene Derivative, Monomalic Acid C₆₀ on Nitric Oxide-Dependent Relaxation of Aortic Smooth Muscle, *Gen. Pharmac.*, Vol.29, No.3, 1997, pp. 345-351.
- [13] Wolff D. J., Mialkowski K., Richardson C. F. and Wilson S. R., C₆₀-Fullerene Monomalonate Adducts Selectively Inactivate Neuronal Nitric Oxide Synthase by Uncoupling the Formation of Reactive Oxygen Intermediates from Nitric Oxide Production, *Biochemistry*, Vol.40, No.1, 2001, pp. 37-45.
- [14] Ueng T.H., Kang J.J., Wang H.W. and Chiang L.Y., Inhibition of Drug-Metabolizing Enzymes in Mouse Liver by a Water Soluble Fullerene C₆₀, *Fullerene Science and Technology*, Vol.7, No.4, 1999, pp. 681-694.
- [15] Dugan L.L., Gabrielsen J.K., Yu S.P., Lin T.S. and Choi D.W., Buckminsterfullerenol Free Radical Scavengers Reduce Excitotoxic and Apoptotic Death of Cultured Cortical Neurons, *Neurobiology of Disease*, Vol.3, 1996, pp. 129-135.
- [16] Fumelli C., Marconi A., Salvioli S., Straface E., Malorni W., Offidani A. M., Pelliccari R., Schettini G., Giannetti A., Monti D. and Pincelli C., Carboxyfullerenes Protect Human Keratinocytes from Ultraviolet-B-Induced Apoptosis, *J. Invest. Dermatol.*, Vol.115, No.5, 2000, pp. 835-41.
- [17] Bisaglia M., Natalini B., Pellicciari R., Straface E., Malorni W., Monti D., Franceschi C. and Schettini G., C₃-Fullerene-tris-Methanodicarboxylic Acid Protects Cerebellar Granule Cells from Apoptosis, *Journal of Neurochemistry*, Vol.74, No.3, 2000, pp. 1197-1204.
- [18] Bosi S., Da Ros T., Castellano S., Banfi E. and Prato M., Antimycobacterial Activity of Ionic Fullerene Derivatives, *Bioorg. Med Chem. Lett.*, Vol.10, No.10, 2000, pp. 1043-1045.
- [19] Hirayama J., Abe H., Kamo N., Shinbo T., Ohnishi-Yamada Y., Kurosawa S., Ikebuchi K and Sekeguchi S., Photoinactivation of Vesicular Stomatitis Virus with Fullerene Conjugated with Methoxy Polyethylene Glycol Amine, *Biology and Pharmacology Bulletin*, Vol.22, No.10, 1999, pp. 1106-1109.
- [20] Da Ros T. and Prato M., Medicinal Chemistry with Fullerenes and Fullerene Derivative, *Chemical Communications*, 1999, pp. 663-669.
- [21] Dugan L.L., Turetsky D.M., Du C., Lobner D., Wheeler M., Almlí C.R., Shen C.K.F., Luh T.Y., Choi D.W. and Lin T.S., Carboxyfullerenes As Neuroprotective Agents, *Proc. Natl. Acad. Sci. USA*, Neurobiology, Vol.94, 1997, pp. 9434-9439.

- [22] Huang S. S., Tsai S. K., Chih C. L., Chiang L. Y., Hsieh H. M., Teng C. M. and Tsai M. C., Neuroprotective Effect of Hexasulfobutylated C₆₀ on Rats Subjected to Focal Cerebral Ischemia, *Free. Radic. Biol. Med.*, Vol.30, No.6, 2001, pp. 643-649.
- [23] Lin A.M.Y., Chyi B.Y., Wang S.D., Yu H.H., Kanakamma P.P., Luh T.Y., Chou C.K. and Ho L.T., Carboxyfullerene Prevents Iron-Induced Oxidative Stress in Rat Brain, *Journal of Neurochemistry*, Vol.72, No.4, 1999, pp. 1634-1640.
- [24] Jin H., Chen W.Q., Tang X.W., Chiang L.Y., Yang C.Y., Schloss J.V. and Wu J.Y., Polyhydroxylated C₆₀, Fullerenols, as Glutamate Receptor Antagonists and Neuroprotective Agents, *Journal of Neuroscience Research*, Vol.62, 2000, pp. 600-607.
- [25] Tsai M.C., Chen Y.H. and Chiang L.Y., Polyhydroxylated C₆₀, Fullerenols, a Novel Free-Radical Trapper Prevented Hydrogen Peroxide- and Cumene Hydroperoxide-elicited Changes in Rat Hippocampus In-vitro, *J. Pharm. Pharmacol.*, Vol.49, 1997, pp. 438-445.
- [26] Chueh S.C., Lai M.K., Lee M.S., Chiang L.Y., Ho T.I. and Chen S.C., Decrease of Free Radical Level in Organ Perfusate by a Novel Water-Soluble Carbon-Sixty-, Hexa(sulfobutyl)-fullerenes, *Transplantation Proceedings*, Vol.31, 1999, pp. 1976-1977.
- [27] Kamat J. P., Devasagayam T. P., Priyadarsini K. I. And Mohan H., Reactive Oxygen Species Mediated Membrane Damage Induced by Fullerene Derivatives and its Possible Biological Implications, *Toxicology.*, Vol.155, No.1-3, 2000, pp. 55-61.
- [28] Lai Y. L., Chiou W. Y., Lu F. J. and Chiang L. Y., Roles of Oxygen Radicals and Elastase in Citric Acid-Induced Airway Constriction of Guinea-Pigs, *British Journal of Pharmacology.*, Vol.126, No.3, 1999, pp. 778-784.
- [29] Lu L.H., Lee Y.T., Chen H.W., Chiang L.Y. and Huang H.C., The Possible Mechanisms of the Antiproliferative Effect of Fullerenol, Polyhydroxylated C₆₀, on Vascular Smooth Muscle Cells, *British Journal of Pharmacology*, Vol.123, 1998, pp. 1097-1102.
- [30] Ueng T.-H., Kang J.-J., Wang H.-W., Cheng Y.-W. and Chiang L.Y., Suppression of Microsomal Cytochrome P450-Dependent Monooxygenases and Mitochondrial Oxidative Phosphorylation by Fullerenol, a polyhydroxylated Fullerene C₆₀, *Toxicology Letters*, Vol.93, 1997, pp. 29-37.
- [31] Hsu H. C., Chiang Y. Y., Chen W. J. and Lee Y. T., Water-Soluble Hexasulfobutyl [60] Fullerene Inhibits Plasma Lipid Peroxidation by Direct Association with Lipoproteins, *J. Cardiovasc. Pharmacol.*, Vol.36, No.4, 2000, pp. 423-427.
- [32] Huang H. M., Ou H. C., Hsieh S. J. and Chiang L. Y., Blockade of Amyloid Beta Peptide-Induced Cytosolic Free Calcium by Fullerenol-1, Carboxylate C₆₀ in PC12 Cells, *Life Sci.*, Vol.66, No.16, 2000, pp. 1525-1533.
- [33] Lee Y. T., Chiang L. Y., Chen W. J. and Hsu H. C., Water-Soluble Hexasulfobutyl [60] Fullerene Inhibit Low-Density Lipoprotein Oxidation in Aqueous and Lipophilic Phases, *Proc. Soc. Exp. Biol. Med.*, Vol.224, No.2, 2000, pp. 69-75.
- [34] Simiæ-Krstiæ J., Effects of C₆₀(OH)₂₄ on Microtubule Assembly, *Archive of Oncology.*, Vol.5, No.3, 1997, pp. 143-145.
- [35] Bogdanoviæ G., Raletia-Savia J., and Markoviæ N., In Vitro Assays for Antitumor-Drug Screening on Human Tumor Cell Lines: Dye Exclusion Test and Colorimetric Cytotoxicity Assay, *Archive of Oncology*, Vol.2, No.4, 1994, pp. 181-184.