Protein Folding by Hydration Aided Search of Minimum Energy

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Abstract: The main result of the computational experiments made in this work is that fitness landscape smoothing has a significant effect on search efficiency. Our test case is related to the famous Levinthal's paradox i.e. how a protein molecule finds its global energy minimum. The energy surface is smoothed by a simple hydration model, which also results in longer range interactions. Hydration also brings more specific correlation between free energy and conformational compactness when compared to the popular lattice models thus aiding conformational search in a way that is not only computationally but also physically sound. Results were compared to those gotten using both the basic local HP model and global Coulombic type models. As a conclusion we can state that based on empirical observations our hydration model implies correlation between energy and compactness, which is beneficial for folding.

Key-Words: fitness landscape, optimisation, protein folding, search

1 Introduction

A protein molecule is a chain of amino acids linked together by peptide bonds i.e. it is an organic heteropolymer. The role of globular proteins in cells is mainly to function as highly specific and efficient chemicals e.g. enzymes and antibodies of the immune system of vertebrates. About half of the proteins of Escherichia coli consist of thousands of enzymes [1]. The rate enhancements produced by enzymes can be extremely high, in excess of 10^{10} [2].

The three-dimensional (native) conformation of protein is called *tertiary structure*. It is widely assumed that the globular shape of globular proteins is due to hydrophobic interactions of the nonpolar atoms tending to create a hydrophobic core covered by polar residues in contact with the solvent (water).

Typically a globular protein consists of 1,000–20,000 atoms and has a diameter of about 35-100Å. In small proteins about half of the atoms are located at the surface while in larger pro-

teins the number falls below 20 percent. Especially in many small proteins disulfide bridges largely contribute to the stability by binding distant parts of the protein together.

Levinthal's paradox: "If a protein is to find its functional conformation by wandering randomly through conformational space, in excess of 10^{50} years would be required for folding." [3, 4, 5, 6]

The number of protein conformations is approximately 8^n , where *n* is the number of residues i.e. each residue has on the average 8 possible conformations. It has been estimated that the number of proteins that could possibly have existed during evolution on Earth is somewhere between 10^{40} and 10^{50} [7, 8].

2 Lattice models

Formally a folding lattice model of a protein can be seen as a finite set V of configurations, an energy function $f: V \to I\!\!R$, and the concept of neighbourhood between the configurations, which means that V is actually a vertex set of a graph Γ , the configuration space, and edge set E defined by the neighbourhood relation. [9] One of the most reduced protein models is the HP model, in which the chain consists of only two types of residues, hydrophobic (H) and polar (P) on a regular 2D or 3D grid, which can further be rectangular or hexagonal. Eaton E. Lattman, Klaus M. Fiebig and Ken A. Dill define H amino acids to be Ala, Ile, Leu, Met, Phe, Trp, Tyr, Val, and Cys while the rest they define to be polar (P) [10]. After this definition every protein sequence can be mapped into a corresponding sequence in HP model space.

The Hamiltonian H of the simple HP model can be written in the form

$$H(\vec{p}, \vec{s}) = -\vec{p} \cdot \vec{s} = \frac{1}{2}(|\vec{s} - \vec{p}|^2 - \vec{p}^2 - \vec{s}^2),$$

where \vec{p} is a vector telling whether the *i*th residue is hydrophobic (1) or not (0) and \vec{s} is a vector telling whether the *i*th site is in the core (1) or on the surface (0). Hence only local interactions are explicitly included in the Hamiltonian, while the longer range interactions only present themselves in the form of steric constraints i.e. as the self avoiding walk (SAW) condition.

The simple HP model is extremely simple. There is no doubt that it lacks many vital features of real proteins. Luckily it is quite easy to make it somewhat more general. In this work we have generalised it by a simple hydration model.

2.1 Fitness landscapes

There are several eminent similarities among the fitness landscapes of complex systems, like protein evolution, spin glasses and satisfiability problems. There are also some differences in the dynamics, which have a profound effect on the long-term evolution with respect to local minima. The problem is that the evolution, or optimisation, easily stops at local extreme. The more rugged landscape the more local extremes and the shorter the average walk to the nearest extremum. While noise, thermal or artificial, is able to help to escape local extremes, it may take too long in practise to use this annealing, real or simulated, scheme when solving complex problems. The situation with protein evolution is totally different. The fitness landscape is rugged, but there is usually always some freedom to move around the local extremal

point. This is because the topology of the fitness landscape: because of *neutral mutations* and the high dimensionality of the sequence space, vast number of sequence combinations, and the short distances between different points, which are actually caused by the high dimensionality.

3 Search method

The key with respect to search efficiency is to combine stochastic and deterministic approaches and in addition to utilise the results already evaluated i.e. to benefit computer memory facility by concentrating search to the neighbourhood of already encountered good trials. This schema is depicted in figure 1: search is done under stochastic control, which inputs best trials from memory to a routine performing local deterministic search. At the beginning of processing the pool of trials is filled with default starting conformation.

The random search procedure is modified so that it keeps the best solutions in memory and tries to find better trials by updating the known best trials. This kind of population based approach is used e.g. in *simulated annealing* and genetic algorithms.



Figure 1: A hybrid conformation search approach.

3.1 Conformation optimisation

Conformation trials are saved to a population where they are after a random number of steps retrieved by a subroutine called loadConformation. The population is initialised by the starting conformation, which is usually the most elongated one. The starting conformation is set elsewhere in the program—usually explicitly supplied by the user. Trials, which are better than the average of the trials, are saved to the population. Corner and crankshaft moves together with totally random point mutation of the conformation are used to find better solutions.

4 Hydration in a lattice model

Hydration can be seen as a process aiding protein folding in several ways. Firstly it mediates the interaction between hydrophobic residues over distances considerable in atomic scale. Secondly hydration inevitably filters, both temporally and especially spatially, interaction potentials. Both phenomena effectively smooth the otherwise very rugged free energy landscape, and thus make the folding process faster. In a way the hydration layer can also be seen to act much as a lubricant between rough surfaces, which reduces friction.

One type of global optimisation method is based on smoothing the object function f(r) by replacing it by a transformed function f(r, t), where t is a control parameter that determines the extent of smoothing. The parameter t is initially set to a large value and then slowly reduced until t = 0, at which point the object function is "in focus"¹ and the global minimum is likely to be revealed. This kind of smoothing algorithm is actually a deterministic analog of the stochastic simulated annealing. Smoothing is also what the next represented method based on a simple hydration model of protein does.

In a conventional lattice model only interactions between a residue and its six nearest neighbours are taken into account when evaluating the free energy. Here we will simulate hydration by interactions depending on a larger neighbourhood having radius of r_H lattice units.

Let us consider the way of evaluating hydration interaction between residues i and j. We have the following principal cases for the distance D(i, j) between residues i and j:

 i = j ± 1 i.e. the bonded case, which will be omitted in our energy calculations,

- $D(i, j) \leq r_H$ i.e. the distance D(i, j) between residues *i* and *j* is less or equal to the radius of the environment and thus we have to evaluate the energy taking also hydration into account, and
- $D(i, j) > r_H$ i.e. the interaction between residues is omitted due to the long distance D(i, j).

The solution adopted here is to simulate hydration iteratively much like running a 3D lattice automaton [11]. In practise the solvent type is replaced by a set of solvent unit cubes having a different hydration degree or level. The number of iteration rounds needed is proportional to r_H [12].

The total free energy due to hydration E_h is evaluated by the formula

$$E_h = -\sum_{i \in NN} h_i,$$

where h_i is the hydration level of voxel *i* and *NN* is the set of voxels surrounding (nearest neighbours) the protein model.

НРРНРНРНРН	$12 \mathrm{mer}$
НРРННРРННРРН	16mer
НРРРРННННРРНРНРНННРНРРННРРН	$27\mathrm{mer}$
РНРРРРРНРРРРНРРРРНРРРРН	27 merPivot
РНРРНРРНРРНРРНРРНРРНРРНР	$27\mathrm{merRep}$
НРРННРРННРРННРРННРРННРРННРРН	$32 \mathrm{mer}$

Table 1: Test sequences

5 Results

The above hydration model was tested using several sequences and hydration parameter combinations. Some of the test sequences can be seen in table 1. The 27merPivot sequence is the least complex of the three sequences that are 27 residues long. It takes about 40,000 local steps to find its native conformation (see table 2). The dependence on r_H does not seem to be high. This is understandable because the simple structure of this sequence supposedly results in a relatively smooth free energy landscape. The relatively smooth landscape is apparently also the reason for the approximately as fast folding

 $^{^1{\}rm smoothing}$ is equivalent to defocusing in optical systems

for the Coulombic type interaction. Folding using the basic HP model is about 30 times slower. Obviously the HP model is spending most of its time to find a needle in a haystack, which is known to be an extremely time consuming search task.

Hydration	interaction
IIYUI ation	mueracuion.

	w_{HH}		Q1	median	Q3		N
	0.5		22,976	$41,\!861$	71,518	1	60
	1.0		$23,\!622$	$42,\!507$	73,734	1	60
	r_H		Q1	median	Q3		N
	2		33,478	$53,\!797$	104,598		80
	3		20,832	$36,\!671$	58,058		80
	4		24,052	$40,\!682$	75,003		80
	5		19,716	38,169	70,330		80
Γ	Psize		Q1	median	Q3		N
	1		19,158	$33,\!595$	60,830		80
	4		23,956	$44,\!107$	73,336		80
	16		28,528	$43,\!170$	80,916		80
	64		25,293	$46,\!546$	$71,\!623$		80
Basic HP model $(r_H = 0)$:							
			Basic HF	P model $(r_I$	H = 0:		
1	Psize		$\frac{\text{Basic HF}}{Q1}$	<u>model (r</u> median	(H = 0): Q	3	N
1	$\frac{Psize}{1}$	7	$\frac{\text{Basic HF}}{Q1}$ $\frac{43,948}{}$	$\begin{array}{c} \textbf{P} \bmod el \ (r_I) \\ \hline median \\ \hline 1,336,524 \end{array}$	(H = 0): Q time-out	3 it	N 20
1	$\frac{Psize}{1}$	7 9	$\frac{\text{Basic HF}}{Q1}$ $\frac{43,948}{37,276}$	$\frac{1}{336,524}$ model (r_1	$\frac{Q}{\frac{Q}{\text{time-out}}}$	3 it it	N 20 20
1	Psize 1 4 16	7 9 6	$\frac{\text{Basic HF}}{Q1} \\ \hline \\ 43,948 \\ 37,276 \\ 14,626 \\ \hline \\$	$\begin{array}{c} {}^{\bullet} \bmod (r_{I}) \\ \hline \\ 1,336,524 \\ 1,574,986 \\ 1,310,792 \end{array}$	$\frac{Q}{\frac{Q}{\text{time-out}}}$	3 it it	N 20 20 20
1	$\frac{Psize}{1}$ 4 16 64	7 9 6 3	$\frac{\text{Basic HF}}{Q1} \\ \hline 43,948 \\ 37,276 \\ 14,626 \\ 93,936 \\ \hline$	$\begin{array}{c} {}^{\bullet} \bmod el \; (r_{I} \\ \hline median \\ 1,336,524 \\ 1,574,986 \\ 1,310,792 \\ 957,568 \end{array}$	$\frac{Q}{Q}$ time-out time-out time-out time-out	3 it it it	$egin{array}{c} N \\ 20 \\ 20 \\ 20 \\ 20 \\ 20 \end{array}$
1	$\frac{P_{size}}{1}$ $\frac{1}{4}$ $\frac{16}{64}$ All	7 9 6 3 6	$\frac{\text{Basic HF}}{Q1} \\ \frac{Q1}{43,948} \\ 37,276 \\ 14,626 \\ 93,936 \\ 65,128 \\ \end{array}$	$\begin{array}{c} {}^{\mathbf{p}} \bmod el \; (r_{I} \\ \hline {median} \\ 1,336,524 \\ 1,574,986 \\ 1,310,792 \\ 957,568 \\ 1,310,792 \end{array}$	$\begin{array}{c} H = 0): \\ \hline Q \\ \hline time-ou \\ time-ou \\ time-ou \\ time-ou \\ time-ou \end{array}$	3 it it it it	N 20 20 20 20 20 80
1	$\begin{array}{c} Psize \\ 1 \\ 4 \\ 16 \\ 64 \\ \hline All \\ Cou \end{array}$	7 9 6 3 6	$\begin{array}{c} \text{Basic HF} \\ \hline Q1 \\ \hline 43,948 \\ 37,276 \\ 14,626 \\ 93,936 \\ \hline 65,128 \\ \hline \text{nbic type} \end{array}$	$\begin{array}{c} \text{P model } (r_1 \\ \hline \text{median} \\ 1,336,524 \\ 1,574,986 \\ 1,310,792 \\ 957,568 \\ \hline 1,310,792 \\ \text{e interactic} \end{array}$	$\begin{array}{l} H = 0): \\ \hline Q \\ time-out \\ time-out \\ time-out \\ time-out \\ time-out \\ \hline n \ (E \propto 1) \end{array}$	$\frac{3}{1t}$ t t t t r)	N 20 20 20 20 80
1	$\begin{array}{c} Psize \\ \hline 1 \\ 4 \\ 16 \\ 64 \\ \hline All \\ \hline Cou \\ \hline Psiz \end{array}$	7 9 6 3 6 lor <i>e</i>	$\begin{array}{c} \text{Basic HF} \\ \hline Q1 \\ \hline 43,948 \\ 37,276 \\ 14,626 \\ 93,936 \\ \hline 65,128 \\ \hline \text{nbic typ} \\ \hline Q1 \end{array}$	$\begin{array}{c} {\rm model} \ (r_1 \\ {\rm median} \\ 1,336,524 \\ 1,574,986 \\ 1,310,792 \\ 957,568 \\ 1,310,792 \\ {\rm e} \ {\rm interactic} \\ {\rm median} \end{array}$	$\frac{Q}{P} = 0):$ $\frac{Q}{P}$ time-out time-out time-out time-out time-out on (E \pm 1)/Q3	$\frac{3}{1t}$	N 20 20 20 20 80
	$\begin{array}{c} Psize \\ \hline 1 \\ 4 \\ 16 \\ 64 \\ \hline All \\ \hline Cou \\ \hline Psiz \\ \hline \end{array}$	7 9 6 3 6 lor 1	$\begin{array}{c} \text{Basic HF} \\ \hline Q1 \\ 43,948 \\ 37,276 \\ 14,626 \\ 93,936 \\ \hline 65,128 \\ \hline \text{nbic typ} \\ \hline Q1 \\ 19,592 \\ \end{array}$	$\begin{array}{c} {\rm model} \ (r_1 \\ {\rm median} \\ 1,336,524 \\ 1,574,986 \\ 1,310,792 \\ 957,568 \\ 1,310,792 \\ {\rm e} \ {\rm interaction} \\ {\rm median} \\ 38,187 \end{array}$	$\frac{Q}{P} = 0):$ $\frac{Q}{V}$ time-out time-out time-out time-out time-out on (E \le 1/ Q3 50,324	$\frac{3}{\text{it}}$ $\frac{1}{\text{it}}$ $\frac{r}{r}$	
]	$\begin{array}{c} Psize \\ \hline 1 \\ 4 \\ 16 \\ 64 \\ \hline All \\ \hline Cou \\ \hline Psiz \\ \hline \end{array}$	7 9 6 3 6 lor e 1 4	$\begin{array}{c} \text{Basic HF} \\ \hline Q1 \\ 43,948 \\ 37,276 \\ 14,626 \\ 93,936 \\ \hline 65,128 \\ \hline \text{mbic type} \\ \hline Q1 \\ 19,592 \\ 17,812 \\ \end{array}$	$\begin{array}{c} {\rm p} \ {\rm model} \ (r_I \\ {\rm median} \\ 1,336,524 \\ 1,574,986 \\ 1,310,792 \\ 957,568 \\ 1,310,792 \\ e \ {\rm interactic} \\ {\rm median} \\ 38,187 \\ 56,281 \end{array}$	$\frac{Q}{E_{H}} = 0):$ $\frac{Q}{E_{H}}$ time-out time-out time-out time-out time-out time-out time-out 0n (E \le 1/ Q3 50,324 76,922	$\frac{3}{1t}$	N 20 20 20 20 80 :
1	$\begin{array}{c c} Psize \\ 1 \\ 4 \\ 16 \\ 64 \\ \hline All \\ \hline Cou \\ \hline Psiz \\ 1 \end{array}$	7 9 6 3 6 1 4 6	$\begin{array}{c} \text{Basic HF} \\ \hline Q1 \\ 43,948 \\ 37,276 \\ 14,626 \\ 93,936 \\ \hline 65,128 \\ \hline \text{mbic type} \\ \hline Q1 \\ 19,592 \\ 17,812 \\ 17,448 \\ \end{array}$	$\begin{array}{r} {\rm model}\;(r_{I}\\ {\rm median}\\ 1,336,524\\ 1,574,986\\ 1,310,792\\ 957,568\\ 1,310,792\\ {\rm e}\;{\rm interactic}\\ {\rm median}\\ 38,187\\ 56,281\\ 41,673\\ \end{array}$	$\frac{Q}{E_{H}} = 0):$ $\frac{Q}{E_{H}}$ time-out time-out time-out time-out time-out time-out time-out 0n (E \le 1/ Q3 50,324 76,922 55,192	$\frac{3}{1t}$	
	$\begin{array}{c c} Psize \\ 1 \\ 4 \\ 16 \\ 64 \\ \hline \\ Cou \\ \hline \\ Psiz \\ \hline \\ 1 \\ 6 \end{array}$	$ \begin{array}{c} 7\\ 9\\ 6\\ 3\\ 6\\ 1\\ 4\\ 6\\ 4\\ 4 \end{array} $	$\begin{array}{r} \hline \\ \textbf{Basic HF} \\ \hline Q1 \\ \hline 43,948 \\ 37,276 \\ 14,626 \\ 93,936 \\ \hline 65,128 \\ \hline \textbf{mbic typ} \\ \hline Q1 \\ 19,592 \\ 17,812 \\ 17,812 \\ 17,448 \\ 17,105 \\ \end{array}$	$\begin{array}{r} {\rm model} \ (r_{I} \\ {\rm median} \\ 1,336,524 \\ 1,574,986 \\ 1,310,792 \\ 957,568 \\ 1,310,792 \\ {\rm e} \ {\rm interaction} \\ {\rm median} \\ 38,187 \\ 56,281 \\ 41,673 \\ 39,300 \\ \end{array}$	$\frac{Q}{E_{H}} = 0):$ $\frac{Q}{E_{H}}$ time-out time	$\frac{3}{1t}$	

Table 2: 27merPivot: The quartiles (Q1, median, Q3) of the number of local steps n_e needed to reveal the native state vs. r_H , population size (Psize), and weight w_{HH} . For the basic HP model max $(n_e) = 2 \times 10^6$ steps, for the others max $(n_e) = 10^6$ steps. N = number of samples.

5.1 Scaling

The above experiments could be extended in many ways. We could collect more statistics on the reference models. But doing this using the current program version and environment would be quite time consuming and that is why it was not included in this work, but left for further studies, which are needed to fully reveal the characteristics of the proposed hydration model and ways to develop it. In any case our simple hydration model was among the fastest model for each test sequence (c.f. tab. 3). Both the basic HP model and the Coulombic model were found to be considerably slower for some of the test sequences.

sequence	$\operatorname{Hydration}$	Basic HP	Coulombic
12-mer	1	3	2
16-mer	1	3	2
27-mer	2	3	1
$27 \mathrm{merPivot}$	1-2	3	1-2
$27 \mathrm{merRep}$	1	3	2
32-mer	1	2	3

 Table 3: Ranking of models vs. test sequences

5.2 Folding as a landscape dependent algorithm

Figure 3 shows schematically how the correlation between free energy E and conformation compactness K varies during protein folding. When the correlation is strongly negative, as it must be at the beginning and also at the end of the process if a stable native state exists, the protein conformation is relatively fast becoming more compact while losing free energy. Ideally a strong negative correlation means efficient hill climbing to the nearest energy minimum. In case of a complex molecule like protein this unfortunately means a high probability of the search getting stuck to a local minimum i.e. a premature convergence. That is why a macromolecule like protein cannot just fall to its free energy minimum conformation. In this case the shortest way to the goal is not the fastest. It is obvious that for most sequences there exists a critical compactness value K_{c_1} which is the highest value still giving for most conformations an energetically and sterically easy access to the native state. From the computational point of view keeping the ensemble below this value is necessary but not sufficient for efficient folding. From the physics point of view passing this value means a high probability of experiencing a glass transition like phase transition.

If the correlation is highly positive, it means that the protein avoids compact states while it loses free energy i.e. there is a strong tendency to avoid trapping to local energy minima. By definition this zone gives an excellent implementation of backtracking the low energy states. Drift away from solution is sometimes called *deception* and it is usually considered a property making the search for optimum difficult.

The third zone in our schematic figure, corr \approx 0, means that there is no correlation between the free energy and the compactness of the conformation. In practise the protein is performing a pure random search driven by thermal noise.

In order for a protein to fold properly within a finite time scale the folding path must in practise draw away from the hill climbing zone to avoid premature convergence. Once the folding ensemble has passed the critical compactness zone it is more and more difficult to have major reconfiguration changes due to the energy and steric constraints. However, the molecule should be as compact as possible when approaching the deception zone, where it is actually driven more towards less compact conformations than the native state. The price to be paid for these conditions is the slowing down of the search, but to avoid Levinthal's paradox the protein must actually slow down the greedy hill climbing search in order to be able to scan the most promising low energy conformations.

6 Conclusions

The effect of hydration on conformation search was tested by searching the native conformations of several test sequences. In general hydration seems to be beneficial. Without it our algorithm needs considerably more local steps. Similarly it seems to be beneficial to count H-H contacts, but their weight used in the free energy formula does not seem to be so critical. Hence it seems that both local and longer range interactions should be modelled in order to have a realistic fast folding process. Moreover a small population size seems to be better than a large one. This is easy to explain: the bigger the population size the more time it takes to process the best trials. Population based search is beneficial because search without a population

of trials is clearly less efficient than that using a population.

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Figure 2: The median number of local steps n_e vs. sequence length n_s The dotted line shows the least squares estimate $N_0 \times 10^{n_s/n_{10}}$, where $N_0 \approx 140$ and $n_{10} \approx 9.2$. As a comparison the dotted curve is $C \times n_s^{3\frac{2}{3}}$, which is a power-law based estimate. For clarity different model shown shifted.



Figure 3: Interpretation of the protein folding path as search algorithms vs. the correlation of energy and conformation compactness K when varying the relative number of H-H contacts n_{HH}/n_{HH} nat. Bold arrows show the direction for decreasing free energy. Notations: $K_c = \text{critical compactness and } \bigcirc = \text{search ensemble}$.