# A method to find protein coding genes in the yeast genome based on a 3D graphical representation of DNA sequence 

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#### Abstract

We develop a method to find protein coding genes based on a 3D graphical representation of DNA sequence. The method is simple and robust. We illustrate it on the yeast genome and it may be extended to find genes in prokaryotic genomes or eukaryotic genomes with less introns. Three-fold cross-validation tests have demonstrated that the accuracy of the algorithm is better than $96 \%$. Based on this, it is found that the total number of protein coding genes in the yeast genome is $5891 \sim 5920$. Among the ORFs annotated in the MIPS database, those recognized as non-coding by the present algorithm are listed in this paper in detail.


Key-Words: - Gene-finding; Yeast genome; 3D graphical representation of DNA sequence

## 1 Introduction

One of the most critical steps of genome annotation is the process of predicting genes that code for proteins. Generally, there are two algorithmic concepts appropriate to recognize genes [1]: 1) A sequence can be classified as a gene, if it shows significant similarity to a sequence, which was annotated as coding and deposited in a database. 2) A statistical analysis of a sequence may indicate its coding potential. This concept is based on the fact that the distributions of nucleotides in coding and non-coding sequences differ statistically significantly [2, 3].
The budding yeast Saccharomyces cerevisiae is an important model organism for the Human Genome Project. As the first sequenced genome of a eukaryotic organism, S. cerevisiae, much work has been done on this aspect. The number of protein coding genes in the yeast genome was estimated to be 5800-6000 [4-6], however, some researchers believe that the number should be less than 4800[7] or 5579[8]. But the prediction of protein coding genes is still far from being a trivial problem.
In this paper we present a simple gene-finding algorithm based on the 3D graphical representation of DNA sequence proposed in [9]. The algorithm utilizes an angle discriminant method to separate the object (ORFs) into two classes of positives (genes) and negatives (non coding ORFs). This simple gene-finding algorithm can perform quickly and it may be complementary with other existing methods.

## 2 Databases and methods

### 2.1 The database

In this paper, all the $S$.cerevisiae genome DNA sequences are taken from http://pedant.gsf.de/ of the Munich Information Center for Protein Sequences (MIPS) released on October 10, 2001. In the MIPS database, all the ORFs are classified into six classes, which correspond to known proteins, strong similarity to known proteins, similarity or weak similarity to known proteins, similarity to unknown proteins, no similarity and questionable ORFs, respectively. The $1^{\text {st }}, 2^{\text {nd }}, 3^{\text {rd }}, 4^{\text {th }}, 5^{\text {th }}$ and $6^{\text {th }}$ classes include 3410 (18), 229, 820(2), 1003, 516, and 471(8) entries, respectively, where the figures in the parentheses indicate the numbers of ORFs in the mitochondrial genome. The mitochondrial ORFs are excluded here since the mitochondrial genetic code differs from the universal genetic code. So in each of the six classes, $3392,229,818,1003,516$, and 463 ORFs are contained, respectively.

### 2.2 The 3D graphical representation of DNA sequence

The three-dimensional graphical representation of DNA sequences provides a visual inspection of DNA data. Several researchers have proposed different graphical representations of DNA sequence [9,14-16].

Our gene-finding algorithm base on the3D
graphical representation of DNA sequence outlined recently in [9]. We present it briefly as follows. We assign A (adenine), G (guanine), T (thymine), and C (cytosine) to $-\mathrm{x},+\mathrm{x},-\mathrm{y}$, and +y , respectively, while the corresponding curve extend along with z -axes. In detail, let $B=b_{1} b_{2} b_{3} \cdots b_{n}$ be an arbitrary DNA sequence. Then we have a map $\Phi_{1}$, which maps $B$ into a plot set. Explicitly, $\Phi_{1}(B)=\Phi_{1}\left(b_{1}\right) \Phi_{1}\left(b_{2}\right) \Phi_{1}\left(b_{3}\right) \cdots \Phi_{1}\left(b_{n}\right)$
where $\Phi_{1}\left(b_{i}\right)= \begin{cases}(-1,0, i) & \text { if } b_{i}=A \\ (1,0, i) & \text { if } b_{i}=G \\ (0,-1, i) & \text { if } b_{i}=T \\ (0,1, i) & \text { if } b_{i}=C\end{cases}$
Connecting adjacent points, we obtain a 3-D curve. In addition, we have another two maps $\Phi_{2}, \Phi_{3}$, where
$\Phi_{2}\left(b_{i}\right)= \begin{cases}(-1,0, i) & \text { if } b_{i}=A \\ (1,0, i) & \text { if } b_{i}=T \\ (0,-1, i) & \text { if } b_{i}=G \\ (0,1, i) & \text { if } b_{i}=C\end{cases}$
$\Phi_{3}\left(b_{i}\right)= \begin{cases}(-1,0, i) & \text { if } b_{i}=A \\ (1,0, i) & \text { if } b_{i}=C \\ (0,-1, i) & \text { if } b_{i}=T \\ (0,1, i) & \text { if } b_{i}=G\end{cases}$

So, for one DNA sequence there are three curves that represent it.

### 2.3. The gene-finding algorithm

Based on two facts (1) amino acid are encoded by triplets of nucleotides of DNA and (2) each nucleotide base does not appear with equal probability at each codon position, comes a conclusion that both the four base (A,C,G, and T) and the three positions are likely to be related with the genetic code $[10,11]$.The curve for the subsequence in an ORF with bases at positions $1,4,7 \cdots$, forms a phase-specific curve. We call this the phase-1 curve. Similarly, the curves with bases at positions $2,5,8 \cdots$, and $3,6,9 \cdots$, are called the phase- 2 and phase- 3 curve, respectively. For an ORF sequence, the phase-1, phase-2, and phase-3 curves describe the distributions of bases at first, second, and third codon positions, respectively. For each phase-specific subsequence, there are three maps $\Phi_{1}, \Phi_{2}, \Phi_{3}$, as for the ordinary DNA sequence. The coordinates of the ith point of phase-j $(\mathrm{j}=1,2,3$ ) curve under the map of $\Phi_{k}(\mathrm{k}=1,2,3)$ are denoted by ( $\left.x_{i, j}^{k}, y_{i, j}^{k}, z_{i, j}^{k}\right)$.

We define

$$
\begin{gathered}
v_{1, j}=\frac{\sum_{i=1}^{N} x_{i, j}^{1}}{z_{N, j}^{1}}, v_{2, j}=\frac{\sum_{i=1}^{N} y_{i, j}^{1}}{z_{N, j}^{1}}, \\
v_{3, j}=\frac{\sum_{i=1}^{N} x_{i, j}^{2}}{z_{N, j}^{2}}, v_{4, j}=\frac{\sum_{i=1}^{N} y_{i, j}^{2}}{z_{N, j}^{2}}, \\
v_{5, j}=\frac{\sum_{i=1}^{N} x_{i, j}^{3}}{z_{N, j}^{3}}, v_{6, j}=\frac{\sum_{i=1}^{N} y_{i, j}^{3}}{z_{N, j}^{3}}, \\
v_{7, j}=\bar{a}^{2}+\bar{c}^{2}+\bar{g}^{2}+\bar{t}^{2}, \text { where } \\
\bar{a}, \bar{c}, \bar{g},
\end{gathered}
$$

and $\bar{t}$ are the average occurrence frequencies of bases A, C, G, and T in the DNA subsequence studied. That is, $\bar{a}=A_{N} / N, \bar{c}=C_{N} / N, \bar{g}=G_{N} / N$, $\bar{t}=T_{N} / N$, where $A_{N}, C_{N}, G_{N}$, and $T_{N}$ are the occurrence numbers of bases $\mathrm{A}, \mathrm{C}, \mathrm{G}$, and T , respectively, in the subsequences, and N is the total length of the subsequence studied. The variable $v_{7, j}$ was found to be a useful statistical quantity for the analysis of DNA sequence [12].

So, for each phase-specific subsequence, there is a seven-dimensional vector $V_{j}=\left(v_{1, j}, v_{2, j}, \cdots, v_{7, j}\right)$, which corresponds to it. We define a 21 -dimensional vector $U=\left(u_{1}, u_{2}, u_{3}, \cdots ; u_{2}\right)$, where

$$
\left\{\begin{array}{lll}
u_{1}=v_{1,1} & u_{2}=v_{2,1} & u_{3}=v_{3,1}  \tag{1}\\
u_{4}=v_{4,1} & u_{5}=v_{5,1} & u_{6}=v_{6,1} \\
u_{7}=v_{7,1} & u_{8}=v_{1,2} & u_{9}=v_{2,2} \\
u_{10}=v_{3,2} & u_{11}=v_{4,2} & u_{12}=v_{5,2} \\
u_{13}=v_{6,2} & u_{14}=v_{7,2} & u_{15}=v_{1,3} \\
u_{16}=v_{2,3} & u_{17}=v_{3,3} & u_{18}=v_{4,3} \\
u_{19}=v_{5,3} & u_{20}=v_{6,3} & u_{21}=v_{7,3}
\end{array}\right.
$$

Therefore, each of the coding ORFs or non-coding DNA sequences is represented by a 21 -dimensional vector.

According to the ergodicity principle, we randomly divide the 3392 genes into two unequal parts, in which the larger part consists of 2000 genes, and the smaller consists of 1392 genes. The former serves as a training set; whereas the latter serves as a test set. Both the training and test sets should be accompanied by the counterparts of negative samples. Considering that the intergenic sequence with length longer than 300 bp , which starts with ATG and ends with one of the stop codons, is
unlikely to be ORF[8,12], we randomly select about 7600 such intergenic sequences from the 16 yeast chromosomes to produce the negative samples. We randomly selected 2000 and 1392 intergenic sequences from the above 7600 sequences, which form the training and test sets of the negative samples, respectively.

The training set of samples (ORFs) is divided into two parts: one includes the positive samples composed of true protein coding genes, the other includes negative samples composed of non-coding DNA sequences. In the positive set the i-th true coding ORF is described by a vector ( $u_{i, 1}^{1}, u_{i, 2}^{1}, \cdots, u_{i, 21}^{1}$ ), where $u_{i, s}^{1}$ are the s-component of the vector ( $s=1,2, \cdots 21$ ). Similarly, in the negative set the i-th non-coding DNA sequences is described by a vector ( $u_{i, 1}^{2}, u_{i, 2}^{2}, u_{i, 3}^{2}, \cdots, u_{i, 21}^{2}$ ), where $u_{i, s}^{2}$ are the s-component of the vector ( $s=1,2, \cdots 21$ ). Suppose the positive and negative sets both include M samples, then we denote the geometric centers of theirs by $\overline{U^{1}}$ and $\overline{U^{2}}$, respectively, where $\overline{U^{1}}=\left(\overline{u_{1}^{1}}, \overline{u_{2}^{1}}, \cdots \overline{u_{21}^{1}}\right), \overline{U^{2}}=\left(\overline{u_{1}^{2}}, \overline{u_{2}^{2}}, \cdots, \overline{u_{21}^{2}}\right) \quad$ and $\overline{u_{s}^{1}}=\frac{1}{M} \sum_{i=1}^{M} u_{i, s}^{1}, \overline{u_{s}^{2}}=\frac{1}{M} \sum_{i=1}^{M} u_{i, s}^{2}(\mathrm{~s}=1,2, \cdots 21)$.

A query ORF is indicated by a 21-dimensional vector $U=\left(u_{1}, u_{2}, \cdots, u_{21}\right)$. To judge whether this ORF is a true protein coding gene or not, calculate the angle $\left\langle U, \overline{U^{1}}\right\rangle$ between $U$ and $\overline{U^{1}}$, and the angle $\left\langle U, \overline{U^{2}}\right\rangle$ between $U$ and $\overline{U^{2}}$, where $\left\langle U, \overline{U^{1}}\right\rangle=\cos ^{-1} \frac{\left(U, \overline{U^{1}}\right)}{|U|\left|\overline{U^{1}}\right|}$ $\left\langle U, \overline{U^{2}}\right\rangle=\cos ^{-1} \frac{\left(U, \overline{U^{2}}\right)}{\left|U \| \overline{U^{2}}\right|}$. A codingness index $\Delta$ is defined as

$$
\begin{equation*}
\Delta=\left\langle U, \overline{U^{2}}\right\rangle-\left\langle U, \overline{U^{1}}\right\rangle+c \tag{2}
\end{equation*}
$$

where $c$ is a constant determined by making false positive rate and false negative rate identical in the training set. If $\Delta>0$, the query ORF is recognized as coding gene, otherwise, if $\Delta<0$, the ORF or DNA sequence is recognized as a non-coding one.

## 3 Results and discussions

### 3.1 Criteria for the evaluation of the

## algorithm

For the evaluation of the performance of the algorithm, we have to discuss the definitions of sensitivity, specificity and selectivity. Denoted by TP the number of coding ORFs that have been correctly predicted as coding, and FN the number of coding ORFs that have been predicted as non-coding. Let TN denote the number of non-coding sequences that have been predicted as non-coding and FP denote the number of non-coding sequences that have been predicted as coding. Then we can define the following term:

$$
\begin{aligned}
& S_{p}(\text { specificity })=\frac{\mathrm{TN}}{\mathrm{TN}+\mathrm{FP}} \\
& \mathrm{~S}_{\mathrm{n}}(\text { sensitivity })=\frac{\mathrm{TP}}{\mathrm{TP}+\mathrm{FN}} \\
& \mathrm{~S}_{1}(\text { selectivity })=\frac{\mathrm{TP}}{\mathrm{TP}+\mathrm{FP}}
\end{aligned}
$$

That is, $S_{n}$ is the proportion of coding ORFs that have been correctly predicted as coding, $S_{p}$ is the proportion of non-coding sequences that have been correctly predicted as non-coding, and $S_{l}$ is the fraction of correctly predicted positive cases among all cases predicted as positive.

The accuracy is defined as the average of $S_{n}$ and $S_{p}$. The definition of accuracy is the same as in [8,12,13]: $A C=\left(S_{n}+S_{p}\right) / 2$

Table 1. The accuracy of the algorithm for three different test sets

| Test set | 1 | 2 | 3 |
| :--- | :--- | :--- | :--- |
| Sensitivity(\%) | 0.974119 | 0.974108 | 0.976276 |
| Specificity(\%) | 0.957585 | 0.950395 | 0.961179 |
| Accuracy(\%) | 0.965852 | 0.9622515 | 0.968728 |

### 3.2 Self-consistency and cross-validation tests

To test the new algorithm, the resubstitution and cross-validation tests are performed. In the version of MIPS database, released on October 10, 2001, the ORFs were classified into six classes, in which the first class consists of 3410 entries corresponding to the known proteins. Excluding the protein coding genes from the mitochondria, 3392 protein genes of the first class residing at the 16 yeast chromosomes remain. The mitochondrial genes are excluded from the present study because the mitochondrial genetic code differs from the universal genetic code.
Using the sequences in the training set, the average vectors $\overline{U^{1}}, \overline{U^{2}}$ and the parameter c are determined. Using these quantities, the accuracy of gene-finding
algorithm in the training and test sets is calculated, which reflects the self-consistency and extrapolating effectiveness of the algorithm. The division of 3392 ORFs into two parts (2000 and 1392) is randomly. Repeating the above random division procedure three times, we have performed three resubstitution and cross-validation tests. In each case, the constant c is determined by making the false positive rate and false negative rate identical in the resubstitution test. The results of the cross-validation test is always greater than $96 \%$, which is higher than that reported in $[8,12$ ] and is comparable to that obtained in [13], however, this method is much faster than the method utilized in [13]. In table 1, the sensitivity, specificity and accuracy of each test are listed.

### 3.3 Apply the algorithm to recognize yeast genes

After performing the resubstitution and cross-validation tests, the 2000 and 1392 positive samples (true genes) are then merged. The 3392 negative samples are selected randomly from the 7600 intergenic sequences mentioned above. These 3392 positive and 3392 negative samples form a new training set. The vectors $\overline{U^{1}}, \overline{U^{2}}$, and the parameter c are obtained.
$\overline{U^{1}}=(0.174627,0.081876,0.037818,-0.054932$, $0.276077,0.119695,0.136808,0.130326,-0.133519$,
0.209294, -0.054551, 0.282182, 0.075775, $-0.078968, \quad 0.087635,-0.131975,0.097351$, $-0.122259, \quad 0.271437,-0.034624,-0.009716)$, $\overline{U^{2}}=(0.141616,-0.144077,0.141884,-0.143808$, $0.275982,-0.002193,-0.000269, \quad 0.146585$, $-0.138505, \quad 0.144128,-0.140963,0.275980$, $0.005623,0.002458,0.145024,-0.127824,0.136119$, $-0.136730, \quad 0.274484,0.008294,0.008905)$, c=0.068875

We then apply the vectors $\overline{U^{1}}, \overline{U^{2}}$, and c listed above to recognizing genes in the ORFs of the $2^{\text {nd }}-6{ }^{\text {th }}$ classes in the MIPS database. For each ORF calculate the vector $U=\left(u_{1}, u_{2}, \cdots, u_{21}\right)$, where $u_{1}, \cdots u_{21}$ are defined in Eq. (1). Based on the vectors $U, \overline{U^{1}}, \overline{U^{2}}$, and the parameter c , calculate the codingness index $\Delta$ using Eq. (2). If $\Delta>0$, the query ORF is recognized as a coding gene, if $\Delta<0$, the ORF or DNA sequence is recognized as a non-coding one. According to the MIPS database, there are 229, $818,1003,516$, and 463 entries of the $2^{\text {nd }}-6^{\text {th }}$ classes in the yeast genome. Consequently, there are 7, 49, 118, 113, 300 entries in the five classes that are recognized as non-coding ORFs. The detailed results are listed in Table 2-6.

Table 2. The 7 ORFs of the $2^{\text {nd }}$ class (strong similarity to known protein) in the MIPS database, which are recognized as non-coding

| $y b r 210 w$ | $y m r 040 w$ | $y e l 004 w$ | ylr046c | yar061w | yll051c | ypl141c |
| :--- | :--- | :--- | :--- | :--- | :--- | :--- |

Table 3. The 49 ORFs of the $3^{\text {rd }}$ class (similarity or weak similarity to known protein) in the MIPS database, which are recognized as non-coding

| ydl199c | yfl040w | yhr130c | yil040w | yjr136c | ylr064w | ylr311c |
| :--- | :--- | :--- | :--- | :--- | :--- | :--- |
| ymr088c | yor053w | yor286w | ybl089w | ybr293w | ydr249c | yer097w |
| yfr057w | ygl160w | yhr181w | yj1091c | yjl193w | ylr365w | ymr221c |
| ymr306w | ynl109w | yol163w | yol079w | ycr001w | ydl206w | ydr119w |
| ydr307w | ydr413c | yel045c | yer113c | yll005c | ylr050c | ylr184w |
| ymr245w | yol107w | yor350c | ykl037w | yal066w | ydr319c | ygr101w |
| ykr030w | ylr283w | ydr115w | ydr366c | ygl104c | ygr284c | yil025c |

Table 4. The 118 ORFs of the $4^{\text {th }}$ class (similarity to unknown protein) in the MIPS database, which are recognized as non-coding

| yar060c | ybr099c | ybr147w | ycr038w-a | ydl240c-a | ydr210w | yer079c-a |
| :--- | :--- | :--- | :--- | :--- | :--- | :--- |
| yer140w | ygl263w | yhl034w-a | yhl045w | yil090w | yir040c | yjr162c |
| ykl225w | ylr149c-a | ylr161w | ylr414c | yml007c-a | ymr010w | ynl156c |
| ynr077c | yol048c | ybl049w | ycl002c | ycl065w | ydr504c | yfr012w |
| ygl041c | ygr016w | yhr017w | ykl106c-a | ymr013w-a | ymr119w | yol002c |
| yol159c-a | yor044w | yor365c | ypl264c | ypr016w-a | yar068w | ybr103c-a |
| ydl027c | ydr084c | ydr438w | ydr492w | yfl015c | yfl062w | yfl068w |
| ygl010w | ygl084c | ygr293c | yhl041w | yhr069c-a | ykl219w | ykr051w |


| yll065w | ylr023c | yml047c | yml132w | ymr326c | ynl326c | yol003c |
| :--- | :--- | :--- | :--- | :--- | :--- | :--- |
| yol162w | ypr071w | yal018c | yal047w-a | ybl029c-a | ybl108w | ybr004c |
| ybr168w | ybr300c | ycr097w-a | ydl185c-a | ydl248w | ydr525w-a | yel053w-a |
| yhl042w | yhr212c | yil174w | yj1097w | ykl223w | ylr156w | ynl067w-a |
| ypr074w-a | ybl109w | ybr191w-a | ybr302c | ycr102w-a | ydl054c | ydl114w-a |
| ydl159w-a | ydr126w | ydr367w | yel033w | yel067c | ygl260w | ygr149w |
| ygr295c | yhl044w | yhr214w-a | yil029c | yil089w | yil175w | yir030w-a |
| yir044c | yj1052c-a | yjr013w | yjr044c | yjr161c | ykl165c-a | ylr036c |
| ylr159w | ynl336w | yol047c | yol101c | yor314w-a | ypl165c |  |

Table 5. The 113 ORFs of the $5^{\text {th }}$ class (no similarity) in the MIPS database, which are recognized as non-coding.

| yar047c | ycl056c | ydr042c | ydr524w-a | yel010w | yfl021c-a | yfr042w |
| :--- | :--- | :--- | :--- | :--- | :--- | :--- |
| ygr168c | ylr111w | ymr151w | ynl324w | yor248w | ypr170w-a | yar053w |
| ybl048w | ybr056w-a | ycl058c | ycr085w | ydl196w | ydr015c | ydr102c |
| ydr274c | ydr396w | yel014c | yel059w | yer135c | ygl188c | yhr139c-a |
| yjl077c | yjl215c | ykr032w | yll030c | ylr112w | yml084w | yml122c |
| ymr057c | ymr320w | yor029w | yor072w | yor314w | yor364w | ypr012w |
| ybl071c | ybr144c | ydr278c | ydr344c | ydr535c | yer066c-a | yer172c-a |
| ygr290w | yhl037c | yhr095w | yir020c-b | yj1028w | yjr157w | ykr073c |
| ylr122c | ylr366w | ylr400w | yml090w | ymr003w | ymr141c | ynl143c |
| ynl211c | yol160w | ypl056c | ypr014c | yal064w | ybr027c | ybr292c |
| ycr022c | ydr024w | ydr179w-a | ydr350c | yer091c-a | ygr026w | ygr291c |
| yil012w | yir020c | yjl136w-a | ykl158w | ylr124w | ylr264c-a | ymr254c |
| ynl146w | ynl174w | ynl303w | yor152c | ypl200w | ypr153w | yar030c |
| yar070c | ycl021w-a | ycr025c | ydr029w | yfl019c | yfr035c | ygl006w-a |
| yhl005c | yjr023c | ykl044w | yll059c | ylr381w | ylr404w | ymr187c |
| ynl150w | ynl179c | yor015w | yor268c | yor392w | ypl041c | ypr064w |
| ypr170c |  |  |  |  |  |  |

Table 6. The 296 ORFs of the $6^{\text {th }}$ class (questionable ORFs) in the MIPS database, which are recognized as non-coding.

| ybl012c | ybl073w | ybr090c | ybr124w | ybr266c | ycr018c-a | ycr087w |
| :--- | :--- | :--- | :--- | :--- | :--- | :--- |
| ydl026w | ydl062w | ydr034c-a | ydr112w | ydr154c | ydr203w | ydr269c |
| ydr355c | ydr431w | ydr467c | ydr526c | yer138w-a | yer181c | yfl032w |
| ygl024w | ygl118c | ygl168w | ygl204c | ygr039w | ygr069w | ygr122c-a |
| ygr176w | yhl006w-a | yhr125w | yil020c-a | yil066w-a | yj1086c | yjl150w |
| yjr018w | ykl030w | ykl136w | ylr101c | ylr198c | ylr322w | ylr428c |
| yml009c-a | yml047w-a | ymr075c-a | ymr316c-a | ynl205c | ynl276c | yor041c |
| yor121c | yor170w | yor225w | yor282w | ypl034w | ypl114w | ypr053c |
| ypr177c | q0143 | yal056c-a | ybl053w | ybl077w | ybl107w-a | ybr224w |
| ycl023c | ycr041w | ydl009c | ydl032w | ydl068w | ydl172c | ydr034w-b |
| ydr114c | ydr157w | ydr360w | yer076w-a | yer107w-a | yer145c-a | yfr036w-a |
| ygl074c | ygl132w | ygl177w | ygr011w | ygr045c | ygr073c | ygr137w |
| ygr182c | ygr259c | yhl019w-a | yil029w-a | yil068w-a | yir023c-a | yjl022w |
| yjr087w | ykl036c | ykl115c | ykl147c | ykl202w | yll020c | ylr123c |
| ylr202c | ylr252w | ylr282c | ylr358c | ylr434c | yml116w-a | ymr086c-a |
| ymr158w-b | ymr290w-a | ynl089c | ynl170w | ynl226w | yol013w-b | yol099c |
| yor135c | yor199w | yor235w | yor300w | yor345c | ypl035c | ypl205c |
| ypr038w | ypr092w | ypr136c | yjr038c | ypr150w | yal026c-a | yal059c-a |
| ybl062w | ybr051w | ybr109w-a | ybr226c | ycl041c | ydl151c | ydl187c |
| ydr048c | ydr133c | ydr220c | ydr290w | ydr401w | ydr442w | ydr509w |
| yel009c-a | yer084w-a | yer148w-a | yfl012w-a | yfr052c-a | ygl042c | ygl088w |


| ygl149w | ygl182c | ygl217c | ygr018c | ygr107w | ygr139w | ygr265w |
| :--- | :--- | :--- | :--- | :--- | :--- | :--- |
| yhl030w-a | yhr063w-a | yhr145c | yil030w-a | yil071w-a | yil163c | yjl032w |
| yjl120w | yjl202c | ykl053w | ykl118w | ylr261c | ylr294c | ylr334c |
| ylr444c | yml012c-a | ymr119w-a | ymr172c-a | ynl013c | ynl105w | ynl171c |
| ynl228w | yol035c | yol106w | yor082c | yor200w | yor309c | ypl044c |
| ypl238c | ypr039w | ypr099c | ypr142c | ypr087w | ypr050c | yal031w-a |
| ybl065w | ybl094c | ybr064w | ybr178w | ydl016c | ydl152w | ydl221w |
| ydr053w | ydr136c | ydr230w | ydr445c | yel018c-a | yer046w-a | yer133w-a |
| yfl013w-a | yfr056c | ygl152c | ygl193c | ygl218w | ygr114c | ygr151c |
| yhl046w-a | yil047c-a | yil100c-a | yjl009w | yj1135w | yjl175w | yjr128w |
| ykl076c | ykr033c | ylr169w | ylr230w | ylr269c | ylr302c | ylr458w |
| yml094c-a | ymr046w-a | ymr304c-a | ynl028w | ynl114c | ynl235c | ynr005c |
| yol037c | yol134c | yor102w | yor146w | yor263c | ypl073c | ypr077c |
| ypr146c | q0092 | yal034c-b | ybl070c | ybr089w | ybr116c | ycr064c |
| ydl050c | ydl094c | ydl158c | ydr008c | ydr149c | ydr199w | ydr241w |
| ydr426c | ydr455c | ydr521w | yel075w-a | yer067c-a | yer087c-a | yer137w-a |
| yer165c-a | ygl109w | ygl165c | ygr025w | ygr064w | ygr115c | ygr228w |
| yhl002c-a | yhr028w-a | yhr071c-a | yil060w | yil115w-a | yir017w-a | yjl015c |
| yjl142c | yjr071w | ykl083w | ykl131w | ylr171w | ylr232w | ylr317w |
| ylr339c | ymr052c-a | ymr153c-a | ymr193c-a | ymr306c-a | ynl120c | ynl198c |
| ynl266w | ynr025c | yol150c | yor169c | yor277c | yor331c | ypl102c |
| ypl185w | ypl261c |  |  |  |  |  |
|  |  |  |  |  |  |  |

Of the entries in above lists, statistically, FN (in list 7) are actually coding. Unfortunately, we cannot identify them at present due to the limited recognition accuracy achieved.

Based on the above result and the sensitivity and specificity, the four quantities TP, TN, FP, and FN can be calculated. Take the $5^{\text {th }}$ class ORFs as an example. The total number of the $5^{\text {th }}$ class ORFs is 516 , in which 113 ones are recognized as non-coding. Assume that both the sensitivity and specificity are equal to $96 \%$. We have a system of linear equations as follows:

$$
\left\{\begin{array}{l}
\mathrm{TP} /(\mathrm{TP}+\mathrm{FN})=0.96 \\
\mathrm{TN} /(\mathrm{TN}+\mathrm{FP})=0.96 \\
\mathrm{TN}+\mathrm{FN}=113 \\
\mathrm{TP}+\mathrm{FN}+\mathrm{TN}+\mathrm{FP}=516
\end{array}\right.
$$

solving the above set of equations, we find $\mathrm{TP} \approx 399$, $\mathrm{TN} \approx 96, \mathrm{FP} \approx 4$, and $\mathrm{FN} \approx 17$. Therefore, the number of real coding ORFs of the $5^{\text {th }}$ class equals to $T P+F N=399+17=416$. Similar calculations for the others are performed. Note that for the $2^{\text {nd }}$ class, the above system has negative solutions: $\mathrm{TP} \approx 222$, $\mathrm{TN} \approx-2, \mathrm{FP} \approx 0, \mathrm{FN} \approx 9$. In this case, we prefer $\mathrm{FN}=7$, $\mathrm{TN}=0$. The results are listed in table 7.

Table 7 The numbers of predicted coding and non-coding ORFs of the $2^{\text {nd }}-6^{\text {th }}$ classes

|  | 2 | 3 | 4 | 5 | 6 |
| :---: | :---: | :---: | :---: | :---: | :---: |
| Total | 229 | 818 | 1003 | 516 | 463 |
| number |  |  |  |  |  |
| of ORFs |  |  |  |  |  |
| TP | 222 | 769 | 882 | 399 | 155 |
| TN | 0 | 16 | 81 | 96 | 289 |
| FP | 0 | 0 | 3 | 4 | 12 |
| FN | 7 | 33 | 37 | 17 | 7 |
| TP+FN | 229 | 802 | 919 | 416 | 162 |
| TN+FP | 0 | 18 | 84 | 100 | 302 |

We estimate the number of protein coding genes in the 16 yeast chromosomes. The total number should be equal to 5920 , the sum of the number of the $1^{\text {st }}$ class and the number of those in the $2^{\text {nd }}-6^{\text {th }}$ classes recognized by the present method. Note that the accuracy is actually greater than $96 \%$, so, this figure should be considered as an upper bound of the number of genes in the yeast genome. Assume that both the sensitivity and specificity are equal to $97 \%$. We also have a system of linear equations. According to the solutions to these system of equations, we can estimate a lower bound of the number of genes in the yeast genome, which is 5891 . The above estimate is based on error analysis, i.e. we have considered the false negative and false positive events in the prediction for each class. So it should be statistically reliable.

## 4 Conclusion

In this paper, a novel gene recognizing method based on a 3D graphical representation of DNA sequence is proposed. As a satisfied result, the successful rates by both self-consistency and cross-validation tests very high and the total number of genes estimated here is $5891 \sim 5920$, coincident with 5800-6000, which is widely accepted. As should be pointed out, to extend the method to more complicated structures, we have not excluded intron-containing genes. The present work is based on an assumption that the unknown genes have the same statistical properties as the known genes. This might not be so in some special cases, for example, for some low-expressed genes. In this case, the results should be referred to with caution.

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